# Specimen Lysis Tubes

For in vitro diagnostic use.

Rx Only

## **Intended Use**

Specimen Lysis Tubes and Specimen Transport Medium (STM) are intended to be used for processing specimens for use with Hologic® assays. Refer to the applicable assay package insert for appropriate Specimen Lysis Tubes and specimen types authorized for use with each assay.

## Materials Provided

#### Available Products

Component	Quantity	Description	Cat. No.
Panther Fusion <sup>®</sup> Specimen Lysis Tube	100 each	1 tube containing 0.71 mL of Specimen Transport Media (STM) with a penetrable cap	PRD-04339
Hologic <sup>®</sup> Specimen Lysis Tube	100 each	1 tube containing 0.71 mL of Specimen Transport Media (STM) with a solid cap	PRD-06554
Hologic <sup>®</sup> Specimen Lysis Tube	1200 each	1 tube containing 0.71 mL of Specimen Transport Media (STM) with a solid cap	PRD-06660
Specimen Transport Medium	1 bottle	Bottle containing 80 mL of Specimen Transport Media (STM)	PRD-04423

#### Materials Required but not Provided

**Note:** Materials available from Hologic have catalog numbers listed, unless otherwise specified.

 Cat. No.

 P1000 pipettor and tips with hydrophobic plugs for the transfer of specimen from the primary collection container to the Specimen Lysis
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 Tube
 Pipet or repeat pipettor capable of 0.78 mL ± 0.07 mL dispense for preparation of Custom Specimen Lysis Tubes only
 —

 Hologic Solid Cap for use with PRD-06554\*, 100 caps per bag \*a single-use cover for the Hologic Specimen Lysis Tube (PRD-06554 only) after testing as part of the uncapped workflow
 PRD-06744

	<u>Cat. No.</u>
Hologic Solid Cap for use with PRD-06660*, 100 caps per bag *a single-use cover for the Hologic Specimen Lysis Tube (PRD-06660 only) after testing as part of the uncapped workflow	PRD-06723
Fisherbrand VersaClosure Tube Closures*, 1000 per pack	02-707
*a single-use tube cover for the Hologic Specimen Lysis Tube (PRD-06554 only) after testing as part of the uncapped workflow	

## **Warnings and Precautions**

Do not apply the transport medium directly to skin or mucous membranes or take internally. For information on any hazards and precautionary statements that may be associated with the specimen lysis tube, refer to the Safety Data Sheet Library at www.hologicsds.com.

#### **Storage Requirements**

Store Specimen Lysis Tubes prior to use at room temperature (15°C to 30°C).

#### **Specimen Performance**

The assay performance characteristics of the specimens are provided in the appropriate Hologic assay package insert. The Hologic assay package inserts may be referenced online at www.hologic.com.

### **Specimen Collection and Handling**

Specimens - Clinical material collected from patient placed in an appropriate transport system.

**Samples** - Represents a more generic term to describe any material for testing on the Panther® or Panther Fusion® System including specimens and specimens transferred into Specimen Lysis Tubes.

**Note:** Handle all specimens as if they contain potentially infectious agents. Use Universal *Precautions.* 

**Note:** Take care to avoid cross-contamination during specimen handling steps. Avoid touching tops of caps of specimens during preparation. Discard used material without passing over open tubes.

Collect specimens according to standard technique. Refer to the assay package insert for the specific sample types compatible with each assay.

#### Specimen Processing using the Panther Fusion Specimen Lysis Tube

A. Prior to testing on the Panther or Panther Fusion system, transfer 500 μL\* of the collected specimen\*\* to a Panther Fusion Specimen Lysis Tube to ensure viral inactivation or specimen lysis.

\***Note:** For Panther Fusion SARS-CoV-2 assay, transfer 250 μL of the LRT specimen and 250 μL of VTM/UTM.

**\*\*Note:** When testing frozen specimen, allow specimen to reach room temperature prior to processing.

B. Refer to the respective assay package insert for information on storage of specimens before and after transferring to the Panther Fusion Specimen Lysis Tube.

#### Specimen Processing using the Hologic Specimen Lysis Tube with Solid Cap

**Note:** Specimen processing using the Hologic Specimen Lysis Tube with solid cap is for use with the Aptima® SARS-CoV-2 assay and the Aptima SARS-CoV-2/Flu assay only.

- A. Uncap the Hologic Specimen Lysis Tube with solid cap and retain the cap.
- B. Prior to testing on the Panther system, transfer 500 μL of the specimen to the Hologic Specimen Lysis Tube with solid cap.
- C. It is recommended to recap the sample tube and gently invert three times to ensure viral inactivation and homogeneous mixture.
- D. To avoid contact with the top of the tube, loosen the cap and place the sample tube into the sample rack.
- E. Remove and discard the cap. To avoid contamination, do not pass the cap over any other sample racks or sample tubes. Inspect the sample tube. If bubbles are present, carefully remove from the sample tube (for example, use the tip of a sterile swab or similar method).

Note: Failure to remove bubbles may affect assay processing and cause invalid results.

F. Place the rack retainer on the sample rack and load the rack into the instrument.

#### Specimen Processing with a Custom Specimen Lysis Tube

**Note:** Specimen processing using the custom Specimen Lysis Tube is for use with the Aptima SARS-CoV-2 assay and the Aptima SARS-CoV-2/Flu assay only.

A. Using a sterile or non-sterile (unused) generic tube made of polypropylene plastic that is 12 mm to 13 mm in outer diameter and 75 mm to 100 mm in height, aliquot 0.78 mL +/- 0.07 mL of bulk STM into the tube using a pipet or repeat pipettor.

**Note:** This step should be conducted in an area where SARS-CoV-2, Flu A, and Flu B specimens are NOT processed

**Note:** If tubes are prepared prior to use, recap the tube and store at 15°C to 30°C until use in specimen processing.

**Note:** When the filled Custom Specimen Lysis Tube is stored closed, if no contaminants were introduced during the filling of the Custom Specimen Lysis Tube, the STM should be stable until the expiration date provided for the STM.

**Note:** There may be an increased risk of contamination when using non-sterile (unused) tubes.

- B. Uncap the custom Specimen Lysis Tube containing STM and retain the cap.
- C. Prior to testing on the Panther system, transfer 500 µL of the specimen to the custom Specimen Lysis Tube containing STM.

- D. It is recommended to recap the sample tube and gently invert three times to ensure viral inactivation and a homogeneous mixture.
- E. To avoid contact with the top of the tube, loosen the cap and place the sample tube into the sample rack.
- F. Remove and discard the cap. To avoid contamination, do not pass the cap over any other sample racks or sample tubes. Inspect the sample tube. If bubbles are present, carefully remove from the sample tube (for example, use the tip of a sterile swab or similar method).

*Note:* Failure to remove bubbles may affect assay processing and cause invalid results.

G. Place the rack retainer on the sample rack and load the rack into the instrument.

#### Sample Storage

- A. Samples on board the Panther system may be archived for additional testing at a later time.
- B. Storing samples after testing
  - 1. Samples that have been assayed should be stored upright in the rack. Refer to the assay package insert for the appropriate storage conditions.
  - 2. The samples should be covered with a new, clean plastic film or foil barrier.
  - 3. If assayed samples need to be frozen or shipped,
    - Capped workflows

Remove the penetrable cap and place a new non-penetrable cap on the specimen tubes. If samples need to be shipped for testing at another facility, recommended temperatures must be maintained. Prior to uncapping, specimen transport tubes must be centrifuged for 5 minutes at 420 Relative Centrifugal Force (RCF) to bring all of the liquid down to the bottom of the tube. Avoid splashing and cross-contamination.

Uncapped workflows

Remove the solid cap and place a new solid cap on the specimen tubes. If samples need to be shipped for testing at another facility, recommended temperatures must be maintained. Prior to uncapping, specimen transport tubes must be centrifuged for 5 minutes at 420 Relative Centrifugal Force (RCF) to bring all of the liquid down to the bottom of the tube. Avoid splashing and cross-contamination.

**Note:** Replacement tube closures and tube plugs should not be used to cover tubes when centrifuging, freezing, or shipping.

### Specimen Transport

Maintain specimen storage conditions as described in the *Specimen Collection and Handling section*.

**Note:** Specimens must be shipped in accordance with applicable national, international, and regional transportation regulations.

## Limitations

Use Specimen Lysis Tubes only with the Hologic assays. Performance has not been established with other products.





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