ThinPrep® Expansion Stain Protocol for the Sakura Tissue-Tek® Prisma® and Prisma® Plus

Autostainers using 95% alcohol fixed slides

Use for staining slides with Small Solution Reservoirs (Sakura P/N 6145). Use this protocol only for staining slides fixed with 95% alcohol.

<u>Caution</u>: Change the Distilled / Instrument Feed water bath at Step 5 (Station 48) after every **NINE** racks of slides. Maintain bath heights to completely cover the slides at full immersion. (Note: Bath fill volume is 280 ml, except for Stations 47 through 52 which are standard, 820 ml baths.)

Required Setup Parameters: Mix (Amplitude = 30 mm, Frequency = 10, Speed = 1), Basket Lifting Speed = 1								
Step	Solution Name	Time	Delay	Mix				
1	Start Station (Reagent or Ethyl alcohol: 70%)	::						
2	Reagent or Ethyl alcohol: 50%	0:01:00	**	on				
3	Distilled or Instrument Feed Water (IFW) ¹	0:01:00	**	on				
4	ThinPrep Nuclear Stain	0:07:00	==	on				
5	Distilled or Instrument Feed Water ¹ (Replace after every 9 racks.)	0:00:10	==	on				
6	ThinPrep Rinse Solution	0:01:00	==	on				
7	Distilled or Instrument Feed Water ¹	0:00:30	==	on				
8	ThinPrep Bluing Solution	0:00:30	==	on				
9	Distilled or Instrument Feed Water ¹	0:00:30	**	on				
10	Reagent or Ethyl alcohol: 50%	0:00:30	**	on				
11	Reagent or Ethyl alcohol: 95%	0:00:30	**	on				
12	ThinPrep Orange G Solution	0:02:00	==	on				
13	Reagent or Ethyl alcohol: 95%	0:00:15	==	on				
14	Reagent or Ethyl alcohol: 95%	0:00:15	==	on				
15	ThinPrep EA Solution	0:04:00	==	on				
16	Reagent or Ethyl alcohol: 95%	0:01:00	==	on				
17	Reagent or Ethyl alcohol: 95%	0:01:00	==	on				
18	Reagent or Ethyl alcohol: 100%	0:00:30	**	on				
19	Reagent or Ethyl alcohol: 100%	0:00:30	**	on				
20	Reagent or Ethyl alcohol: 100%	0:00:30	**	on				
21	Xylene or other Hologic approved clearing agent ²	0:01:00	**	on				
22	Xylene or other Hologic approved clearing agent ²	0:03:00	**	on				
23	End Station (Xylene or other Hologic approved clearing agent 2)	::						
Remove slides to a separate clearing bath then coverslip with the appropriate Hologic-approved mountant. ²								

^{**} Unlimited, == Exact (no delay)

² See ThinPrep Stain User's Manual, Section 1, "STAINING" or contact Hologic for current list of Hologic approved clearing agents and mounting medias.



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¹ Hologic specification for IFW: ≥1.0 megohm-cm Resistivity <u>OR</u> ≤1.0 μSiemens/cm Conductivity (Refer to Clinical and Laboratory Standards Institute (CLSI) document C3-A4, 2006)

Sakura Tissue-Tek Prisma and Prisma Plus Autostainers

For staining slides fixed with 95% alcohol

1	3	5	7 (W4)	8 (W3)	9 (W2)	10 (W1)	13 (D1a)
THINPREP	95%	50%					
ORANGE G	ALCOHOL	ALCOHOL					
12	11	10					
2	4	6	11	12	54	55	14 (D1b)
95%	95%	DISTILLED					11(212)
ALCOHOL	ALCOHOL	WATER					
13	14	9					
15	17	19	21	23	25	27	29 (D2a)
95%	THINPREP	THINPREP					
ALCOHOL	EA	BLUING					
16	15	8					
16	18	20	22	24	26	28	30 (D2b)
95%	100%	DISTILLED					
ALCOHOL 17	ALCOHOL 18	WATER 7					
17	10	/					
31	33	35	37	39	41	43	45*
100%	100%	THINPREP					
ALCOHOL 20	ALCOHOL 19	RINSE 6					
32	34	36 THINPREP	38	40	42	44	46*
XYLENE	XYLENE	NUCLEAR					
21	22	STAIN 4					
53	47 (E1)	48 (E2)	49 (E3)	50 (S3)	51 (S2)	52 (S1)	
53	XYLENE	POST- NUCLEAR	DISTILLED	50%	70%	70%	
	ATLEINE	DISTILLED	WATER	ALCOHOL	ALCOHOL	ALCOHOL	
LINK	23 / End	WATER 5	3	2	1 / Start	1 / Start	
				L			
	E# = Exit St	tation	W# = Running	water S#	# = Start Station	D# = D	Orying Station

^{*} Configuration shown is for Prisma Model 6130 or Prisma Plus Model 6170. In Models 6131 and 6171, Stations 45 & 46 are replaced with heat stations.